EDITORIAL

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Current Status and Perspectives of Phage Therapy*

Terapia fagowa – stan obecny i perspektywy

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A dramatic rise in antibiotic resistance in many clinically significant bacterial species [1] coupled with a shortage of novel classes of antibiotics [2] has created an urgent need for the development of alternative antibacterial agents [3, 4]. One of the most interesting and promising of such therapeutic modalities are currently lytic bacteriophages (phages), i.e. highly specialized viruses that infect and kill solely bacterial cells [4, 5]. These were dis− covered by Frederick Twort in 1915 and, indepen− dently, by Felix d'Herelle in 1917. Although at the beginning of the twentieth century knowledge about bacteriophages was very scant, some researchers even then realized phages' antibacteri− al activity and attempts were made to exploit it for therapeutic purposes, the first clinical trial taking place in Paris in 1919. Thus the history of phage treatment of bacterial infections encompasses an impressive 87 years – almost 30 years more than the antibiotic era. However, the therapeutic and prophylactic trials that were conducted in the early part of the twentieth century were often unsuccess− ful and, accordingly, the first phage researchers failed to inculcate the idea of bacteriophage ther− apy in Western medicine. Basically, the major rea− sons for this failure were inadequate knowledge of phage biology and a poor standard of scientific research. Thus, not surprisingly, interest in the ther− apeutic use of bacteriophages dwindled in the West following the introduction of antibiotics for the treatment of infectious diseases. However, bacte− riophages continued to be used in Eastern Europe,

especially in Poland and the former Soviet Union (Georgia and Russia), the two leading centers being the Institute of Immunology and Experi− mental Therapy (IITD) in Wrocław and the Eliava Institute in Tbilisi [6, 7].

Bacteriophages as Antibacterial Agents

One of the most significant features of bacte− riophages as potential antibacterial agents is their narrow antibacterial range. Essentially, lytic bacte− riophages infect bacteria in a sub−species−specific manner, i.e. they are capable of killing only certain strains within a given bacterial species [3]. For example, the enterococcal phage ENB6, studied with a view to potential therapeutic use, was found to kill 57% of different clinical isolates of VRE tested and to inhibit the growth of an additional 22% of the isolates [8]. On one hand, this feature appears to be advantageous, as phages, unlike antibiotics, can clear pathogenic bacteria without disturbing the balance of the indigenous bacterial microflora [3]. On the other hand, however, a nar− row antibacterial range may be deemed a draw− back, because it requires determining whether the bacteria causing infection in a given patient are sensitive to phages *in vitro* [9]. Alternative approaches include administration of a phage "cocktail", i.e. a mixture of a few different phages, collectively providing a wider antibacterial range

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[9], or the use of a single phage with a broader antibacterial spectrum [10].

With respect to the major mode of the antibac− terial action of bacteriophages, it is rather far more complicated than the mechanisms of action of typ− ical small−molecule antibiotics and is part of the complex interaction between phage virions and the host bacterial cell. Basically, over the course of infection of a susceptible bacterial cell, lytic bac− teriophages divert various essential bacterial meta− bolic pathways (e.g. protein biosynthesis or ATP generation) from their normal functions and direct them towards phage progeny production [11]. A growing body of experimental findings clearly shows that it is indeed direct killing of bacteria by phage virions that forms the basis of the therapeu− tic effect of bacteriophages [8, 12–15]. The other potential mechanism, i.e. the induction of an immune response by some component(s) of the bacteriophage preparation, plays a minor role, if any, in that regard. It was shown in elegant exper− iments that only functional phage particles, i.e. those having the capacity to kill bacteria *in vitro,* were capable of curing mice from a potentially lethal infection, whereas preparations containing functionally inactivated phage failed [8, 13, 14].

A therapeutic strategy was recently reported that apparently may constitute a viable alternative to the use of lytic bacteriophages. This method relies on using a nonlytic filamentous phage (e.g. M13), which basically replicates in the host bacterial cell without killing it. In this case, the virions act as vehicles delivering DNA encoding bactericidal pro− teins, e.g. addiction toxins or restriction endonucle− ases, to bacterial cells. The antibacterial activity of such recombinant phages does not result from phage virions themselves killing bacterial cells, but from the bactericidal activity of the proteins, pro− ducts of genes which were introduced into the bac− teria. The effectiveness of this novel therapeutic approach was shown both *in vitro* and *in vivo* [16].

In view of the growing menace of multi−drug− resistant bacteria, of paramount importance is the fact that the mode of antibacterial action of bacterio− phages is entirely distinct from those employed by traditional antibiotics. This feature seems to render phages particularly useful in the treatment of infec− tions caused by antibiotic−resistant bacteria [17]. Indeed, in experimental studies bacteriophages proved highly effective in that regard, being capable of rescuing mice challenged with a lethal dose of vancomycin−resistant enterococci [8], imipenem− resistant *P. aeruginosa* [13], *E. coli* ESBL [14], as well as methicillin−resistant *S. aureus* [15].

For many years, antibiotics have been, and still are, the unquestionable standard in the treat− ment of bacterial infections. Thus, the obvious

question arises whether the therapeutic efficacy of bacteriophages can exceed that of traditional small−molecule antibacterial agents. This funda− mental issue was addressed in a classical study by Smith and Huggins, who showed that one dose of coliphage was more effective than multiple doses of four different antibiotics in curing mice of a potentially lethal *E. coli* infection. Apparently, the superiority of bacteriophage over antibiotics resulted from the phage−unique capacity for expo− nential growth, a phenomenon dependent upon replication within bacterial cells [12]. Thus, bacte− riophages are the only known class of antibacteri− al agents whose titer (i.e. an equivalent of the con− centration of an antibiotic) grows over the course of treatment, thereby increasing its efficacy.

Contemporary phage therapy has benefited considerably not only from a better understanding of phage biology, but also from substantial progress in molecular biology and chemistry. For example, thanks to major advances in the knowl− edge of bacteriophage life cycles in bacterial cells, it is currently known that only lytic phages may be used for therapeutic purposes, whereas temperate ones should be excluded from treatment (the latter do not kill the host cell upon entering it, but rather integrate their genome into the host chromosome) [9]. Moreover, the genomes of phages to be used in treatment may be sequenced to determine if any of the putative phage proteins have any homologies to potentially deleterious bacterial proteins, including toxins, pathogenicity factors, or antibiotic−resis− tance determinants [18]. Another factor substan− tially contributing to improvement in the safety of phage therapy is the development of novel purifi− cation protocols of crude bacteriophage suspen− sions. At the IITD, for instance, a method was developed that enables one to obtain highly puri− fied preparations of bacteriophages specific to dif− ferent Gram−negative bacterial species. Such preparations formerly contained considerable amounts of endotoxin. It is now possible to obtain preparations contaminated with a mere 0.4–7 EU of endotoxin per milliliter, a value allowing even intravenous administration [19].

The results of well−controlled preclinical studies clearly point to the high efficacy of phage therapy. However, these studies have also revealed that bac− teriophages, as all other classes of antibacterial agents, do have some inherent drawbacks which may considerably diminish their therapeutic effec− tiveness. The importance of a narrow antibacterial range – a classical problem associated with phage therapy – was mentioned above. Another major problem which definitely diminishes the antibacter− ial activity of bacteriophages *in vivo* is their rapid clearance, determined largely by the non−specific

entrapment of phage particles by cells of the reticu− lo−endothelial system (RES) of the liver and spleen. However, it is possible to isolate, relatively easily, phage mutants featuring a considerably prolonged serum half−life. Predictably, such phages exert a more potent antibacterial activity *in vivo* [20]. Another factor that is believed to diminish the ther− apeutic effectiveness of bacteriophages is specific humoral immunity, i.e. the generation of neutralizing anti−phage antibodies. Such antibodies are very like− ly to disturb the interaction between phage virions and bacterial cells [3, 9]. To the best of our knowl− edge, no reliable solution to this problem has been reported as yet. While the immunogenicity of pro− tein pharmaceuticals can be considerably reduced by means of conjugation to polyethylene glycol (PEG) [21], bacteriophages have a far more complicated structure, their particles being made up of many dif− ferent proteins. Thus, the pegylation of phage viri− ons to reduce their immunogenicity is apparently not feasible. Perhaps increasing the dose of bacterio− phage or administering a phage with a different anti− genic specificity following the generation of neutral− izing antibodies could be helpful in this regard.

One of the major problems in antimicrobial ther− apy is the development of resistance. In fact, it is the sharp rise in the prevalence of antibiotic−resistant bacterial strains that has prompted a resurgence of interest in phage therapy in Western medicine over the past years [3, 4]. Thus the question arises whether or not bacteriophages will meet the fate of the traditional, small−molecule antibacterial agents, i.e. an increasing frequency of treatment failures owing to the emergence of multi−resistant bacterial strains. Apparently they will not. First, over the very long co−evolution with their host cells, bacterio− phages have developed some very effective means of dealing with bacteria [22], an example being the activity of the highly evolved endolysins (see below). Accordingly, it is believed that resistance to phages develops ten times more rarely than resis− tance to antibiotics [9]. Furthermore, the develop− ment of resistance can be considerably delayed by using a phage "cocktail" [23]. Importantly, phage− resistant bacterial mutants were found to feature a lowered virulence *in vivo* [12]. Thus we have rea− son to believe that the development of resistance – an apparent and inevitable consequence of using any antibacterial agent – will in fact not have as dramat− ic an effect on phage therapy as it had on antibiotics.

Phage Therapy in Humans

Over the past few years, the history, current sta− tus, and future prospects of phage therapy in humans have been commented and reviewed extensively [5–7, 9, 24–27]. Of special importance are the papers published in top−tier journals, such as The Lancet [28, 29], JAMA [30], Science [31], Nature Biotechnology [4], and Nature Reviews [3, 32], as they reflect the great significance of the topic. As pointed out in Science [31] and in an excellent review in Antimicrobial Agents and Chemotherapy [7], the most detailed series of studies were conduct− ed in Poland using phage preparations developed at the IITD. The major conclusions arising from these studies may be summarized as follows: 1) Bacteriophages are very effective in many different kinds of infections caused by both Gram−positive and Gram−negative antibiotic−resistant bacterial strains, with an overall therapeutic success rate exceeding 85%. 2) Bacteriophages are efficacious in both non−invasive and invasive infections (e.g. sep− ticemia). 3) Phages can be used in both mono− and polyinfections. 4) Bacteriophages can be adminis− tered along with antibiotics, though in this case they are significantly less effective. 5) The treatment is apparently safe, with no side−effects occurring in patients following phage administration [33] (more data are available on our website: http://surfer.iitd. pan.wroc.pl/phages/phages.html). However, it should be kept in mind that these studies were not con− trolled studies, as were practically all of the others regarding phage therapy in humans (several hundred papers in all). Accordingly, they do not meet current rigorous standards for clinical trials and cannot pro− vide ultimate confirmation of either the effectiveness or the safety of phage therapy. Thus one of the great− est challenges to be met in the nearest future is to conduct a controlled clinical trial using a well−char− acterized phage preparation. The first step towards achieving this goal was recently made with the car− rying out of the first well−controlled safety test of phage therapy, during which no side−effects occurred in 15 healthy volunteers following oral administration of T4 coliphage [34].

Other Activities of Bacteriophages

Since the discovery of bacteriophages at the beginning of the twentieth century, their major medical application has been the treatment of bac− terial infections. However, we believe that phages should not be pigeonholed as merely "viruses of bacteria", as they are also capable of exerting other and sometimes unexpected activities, some of which have already been exploited in medicine. For instance, a considerable body of experimental evidence indicates that phages may also be used for the treatment of viral infections [for a review, see Ref. 35]. Basically, there are two major mech−

anisms by which bacteriophages may diminish the infectivity of pathogenic viruses. First, phage nucleic acids can induce the synthesis of interfer− ons, cytokines known to exert a potent anti−viral activity. The other mechanism could be direct competition of phage particles and pathogenic virions for cellular surface receptors. More recent− ly, a third possible mechanism has been reported, i.e. direct binding of viral proteins by bacterio− phage proteins, leading to an inhibition of their natural activity [36]. Moreover, phages can be exploited for the development of anti−viral vac− cines in which phage particles constitute delivery vehicles for either vaccine antigens themselves or their corresponding DNA sequences ("DNA vac− cines") [37]. Although the majority of the relevant findings come from preclinical studies, there are some encouraging data in the literature suggesting that the anti−viral activity of bacteriophages can also be successfully employed in the treatment of viral infections in humans [35].

Another interesting (and potentially very important) activity of bacteriophages is their effect on the immune system. For example, our group showed that at least some phages specific to dif− ferent species of Gram−negative bacteria can exert immunosuppressive activity, an example of such a phage being T4 coliphage, which was found to inhibit human T−cell proliferation, mouse antibody production, as well as NF−kappaB activation *in vitro*. These findings gain special significance in the context of the postulated use of bacteriophages in the treatment of bacterial infections in allograft recipients. In such patients, an immunostimulative effect of phage could accelerate allograft rejection, whereas the immunosuppressive activity could be beneficial. Here it is worth pointing out that T4 was found to actually extend skin allograft sur− vival in mice and to diminish cellular infiltration of the graft [38, 39].

Endolysins

In 2001, a very interesting alternative to the classical phage therapy was reported, i.e. recombi− nant endolysins, or lysins [40]. These are dsDNA bacteriophage−encoded enzymes that are produced in phage−infected bacterial cells during the later stages of the lytic cycle, their major function being the cleavage of peptidoglycan covalent bonds, which results in lysis of the host bacterial cell and ensures the successful release of progeny virions [41]. Over the course of phage infection of a bac− terial cell, endolysin molecules are synthesized in the cytoplasm and reach their substrate, peptido− glycan, from within the cell. The feasibility of using lysins as antibacterial agents results from the fact that they are capable of cleaving peptidogly− can also when applied exogenously (as purified recombinant proteins) to the bacterial cell wall. In this case, their lytic effect is very rapid and potent, especially in Gram−positive bacteria, whereas Gram−negative bacteria are generally considered to be resistant owing to the presence of the outer membrane, which blocks the access of lysin mole− cules to peptidoglycan [41, 42]. Accordingly, one of the biggest challenges endolysin researchers must now face is finding some means of enabling lysin molecules to penetrate through the outer membrane. In fact, some findings indicate that at least some lytic enzymes can also be successful in killing Gram−negative bacteria when acting on them from outside [42].

The most important features of endolysins as potential antibacterial agents include: 1) a very rapid and potent antibacterial activity against Gram−positive bacteria both *in vitro* and *in vivo*, 2) a novel mode of antibacterial action associated with enzymatic cleavage of peptidoglycan cova− lent bonds, 3) the capability to kill bacteria regard− less of their antibiotic sensitivity, 4) a narrow, species−specific antibacterial range, 5) a very low probability of the development of resistance, 6) apparent safety, and 7) relatively easy modifica− tions using genetic engineering. These features clearly set them apart from traditional antibiotics and create a truly novel and unique class of antibacterial agents [42]. Lytic enzymes were orig− inally developed with a view to killing Gram−pos− itive bacteria colonizing mucous membranes. This colonization is of great importance to medicine, as it provides a potential starting point for infection and contributes to the horizontal spread of bacteria within the community. Hence, owing to their rapid killing of bacteria in a basically species−specific manner, lysins provide a unique means of selective prophylaxis of infections without disturbing the balance of the indigenous microflora [43]. In fact, several studies have clearly shown a great capaci− ty of lytic enzymes for killing bacteria colonizing mucous membranes of mice following topical administration [40, 44, 45]. The other potential application of lysins may be the treatment of bac− terial infections, the results of the first relevant studies being very encouraging [46, 47]. Interestingly, it has been shown that antibodies, contrary to expectations, do not neutralize, but rather slightly decrease the antibacterial activity of lytic enzymes *in vivo* [46, 47]. This finding is very important, as it provides an additional argument in favor of the possibility of using lysins for the treat− ment of systemic bacterial infections.

Endolysins and bacteriophages have a few sig−

nificant features in common. These include: a novel (as compared with antibiotics) mode of action, the capability to kill bacteria regardless of their antibiotic−sensitivity, and a narrow antibacte− rial range. On the other hand, there are also some major differences between them, as endolysins, unlike bacteriophages, do not have the capacity for exponential growth and are less likely to be used for the treatment of Gram−negative bacterial infec− tions. However, owing to the lack of reports direct− ly comparing the antibacterial activity of lysins and phages, no general conclusions can be drawn regarding the superiority of either modality.

Over past years we have been witnessing a great resurgence of interest in phage therapy. Bacterio−

phages possess several important features which col− lectively set them clearly apart from traditional antibiotics and render them a unique class of antibacterial agents. Judging from the results of pre− clinical studies and clinical trials conducted hitherto, phage therapy appears to be very effective and safe. However, as impressive as some of these results are, they must be ultimately verified by controlled clini− cal trials. We believe that further research will con− firm both the high effectiveness and the safety of phage therapy and that this therapeutic modality will soon become a widely accepted way of treating infections caused by antibiotic−resistant bacteria – a great challenge of modern medicine.

References

- **[1] Walsh FM, Amyes SGB:** Microbiology and drug resistance mechanisms of fully resistant pathogens. Curr Opin Microbiol 2004, 7, 439–444.
- **[2] Breithaupt H:** The new antibiotics. Nat Biotechnol 1999, 17, 1165–1169.
- **[3] Merril CR, Scholl D, Adhya SL:** The prospect for bacteriophage therapy in Western medicine. Nat Rev Drug Discov 2003, 2, 489–497.
- [4] Thiel K: Old dogma, new tricks -21 st century phage therapy. Nat Biotechnol 2004, 22, 31–36.
- **[5] Lorch A:** Bacteriophages: an alternative to antibiotics? Biotechnol Dev Monitor 1999, 39, 14–17.
- **[6] Debattista J:** Phage therapy: where East meets West. Expert Rev Anti Infect Ther 2004, 2, 815–819.
- **[7] Sulakvelidze A, Alavidze Z, Glenn Morris J Jr.:** Bacteriophage therapy. Antimicrob Agents Chemother 2001, 45, 649–659.
- **[8] Biswas B, Adhya S, Washart P, Paul B, Trostel AN, Powell B, Carlton R, Merril CR:** Bacteriophage therapy rescues mice bacteremic from a clinical isolate of vancomycin−resistant *Enterococcus faecium*. Infect Immun 2002, 70, 204–210.
- **[9] Carlton R:** Phage therapy: past history and future prospects. Arch Immunol Ther Exp 1999, 47, 267–274.
- **[10] Schoolnik GK, Summers WC, Watson JD:** Phage offer a real alternative. Nat Biotechnol 2004, 22, 505–506.
- **[11] Campbell A:** The future of bacteriophage biology. Nat Rev Gen 2003, 4, 471–477.
- **[12] Smith HW, Huggins MB:** Successful treatment of experimental Escherichia coli infections in mice using phage: its general superiority over antibiotics. J Gen Microbiol 1982, 128, 307–318.
- **[13] Wang J, Hu B, Xu M, Yan Q, Liu S, Zhu X, Sun Z, Reed E, Ding L, Gong J, Li QQ, Hu J:** Use of bacterio− phage in the treatment of experimental animal bacteremia from imipenem−resistant *Pseudomonas aeruginosa*. Int J Mol Med 2006, 17, 309–317.
- **[14] Wang J, Hu B, Xu M, Yan Q, Liu S, Zhu X, Sun Z, Tao D, Ding L, Reed E, Gong J, Li QQ, Hu J:** Therapeutic effectiveness of bacteriophages in the rescue of mice with extended spectrum beta−lactamase−producing *Escherichia coli* bacteremia. Int J Mol Med 2006, 17, 347–355.
- **[15] Matsuzaki S, Yasuda M, Nishikawa H, Kuroda M, Ujihara T, Shuin T, Shen Y, Jin Z, Fujimoto S, Nasimuzzaman MD, Wakiguchi H, Shigeyoshi S, Sugiura T, Koda S, Muraoka A, Imai S:** Experimental pro− tection of mice against lethal *Staphylococcus aureus* infection by novel bacteriophage ØMR11. J Inf Dis 2003, 187, 613–624.
- **[16] Westwater C, Kasman LM, Schofield DA, Werner PA, Dolan JW, Schmidt MG, Norris JS:** Use of geneti− cally engineered phage to deliver antimicrobial agents to bacteria: an alternative therapy for treatment of bacteri− al infections. Antimicrob Agents Chemother 2003, 47, 1301–1307.
- **[17] Sulakvelidze A:** Phage therapy: an attractive option for dealing with antibiotic−resistant bacterial infections. Drug Discov Today 2005, 10, 807–809.
- **[18] Carlton RM, Noordman WH, Biswas B, de Meester ED, Loessner MJ:** Bacteriophage P100 for control of Listeria monocytogenes in foods: genome sequence, bioinformatic analyses, oral toxicity study, and application. Reg Toxicol Pharmacol 2005, 43, 301–312.
- **[19] Boratynski J, Syper D, Weber−Dabrowska B, Lusiak−Grzelachowska M, Pozniak G, Gorski A:** Preparation of endotoxin−free bacteriophages. Cell Mol Biol Lett 2004, 9, 253–259.
- **[20] Merril C, Biswas B, Carlton R, Jensen NC, Creed GJ, Zullo S, Adhya S:** Long−circulating bacteriophage as antibacterial agents. Proc Natl Acad Sci USA 1996, 93, 3188–3192.
- **[21] Veronese FM, Pasut G:** PEGylation, successful approach to drug delivery. Drug Discov Today 2005, 10, 1451–1458.
- **[22] Comeau AM, Krisch HM:** War is peace: dispatches from the bacterial and phage killing fields. Curr Opin Microbiol 2005, 8, 488–494.
- **[23] Tanji Y, Shimada T, Fukudomi H, Miyanaga K, Nakai Y, Unno H:** Therapeutic use of phage cocktail for con− trolling Escherichia coli O157:H7 in gastrointestinal tract of mice. J Biosci Bioeng 2005, 100, 280–287.
- **[24] Barrow PA, Soothill JS:** Bacteriophage therapy and prophylaxis: rediscovery and renewed assessment of poten− tial. Trends Microbiol 1997, 5, 268–271.
- **[25] Duckworth DH, Gulig PA:** Bacteriophages. Potential treatment for bacterial infections. Biodrugs 2002, 16, 57–62.
- **[26] Summers WC:** Bacteriophage therapy. Annu Rev Microbiol 2001, 55, 437–451.
- **[27] Alisky J, Iczkowski K, Rapoport A, Troitsky N:** Bacteriophage show promise as antimicrobial agents. J Infect 1998, 36, 5–15.
- **[28] Bradbury J:** My enemy's enemy is my friend. Using phages to fight bacteria. Lancet 2004, 363, 624–625.
- **[29] Parfitt T:** Georgia: an unlikely stronghold for bacteriophage therapy. Lancet 2005, 365, 2166–2167.
- **[30] Thacker PD:** Set a microbe to kill a microbe. Drug resistance renews interest in phage therapy. J Am Med Assoc 2003, 290, 3183–3185.
- **[31] Stone R:** Stalin's forgotten cure. Science 2002, 298, 728–731.
- **[32] Levin BR, Bull JJ:** Population and evolutionary dynamics of phage therapy. Nat Rev Microbiol 2004, 2, 166–173.
- **[33] Weber−Dabrowska B, Mulczyk M, Gorski A:** Bacteriophage therapy of bacterial infections: an update of our Institute's experience. Arch Immunol Ther Exp 2000, 48, 547–551.
- **[34] Bruttin A, Brussow H:** Human volunteers receiving Escherichia coli phage T4 orally: a safety test of phage ther− apy. Antimicrob Agents Chemother 2005, 49, 2874–2878.
- **[35] Miedzybrodzki R, Fortuna W, Weber−Dabrowska B, Gorski A:** Bacterial viruses against viruses pathogenic for man? Virus Res 2005, 110, 1–8.
- **[36] Krichevsky A, Rusnati M, Bugatti A, Waigmann E, Shokat S, Loyter A:** The fd phage and a peptide derived from its p8 coat protein interact with the HIV−1 Tat−NLS and inhibits its biological functions. Antiviral Res 2005, 66, 67–78.
- **[37] Clark JR, March JB:** Bacterial viruses as human vaccines? Expert Rev Vaccines 2004, 3, 463–476.
- **[38] Gorski A, Nowaczyk M, Weber−Dabrowska B, Kniotek M, Boratynski J, Ahmed A, Dabrowska K, Wierzbicki P, Switala−Jelen K, Opolski A:** New insights into the possible role of bacteriophages in transplanta− tion. Transplant Proc 2003, 35, 2372–2373.
- **[39] Gorski A, Kniotek M, Perkowska−Ptasinska A, Mroz A, Przerwa A, Gorczyca W, Dabrowska K, Weber− −Dabrowska B, Nowaczyk M:** Bacteriophages and transplantation tolerance. Transplant Proc 2006 38, 331–333.
- **[40] Nelson D, Loomis L, Fischetti VA:** Prevention and elimination of upper respiratory colonization of mice by group A streptococci by using a bacteriophage lytic enzyme. Proc Natl Acad Sci USA 2001, 98, 4107–4112.
- **[41] Fischetti VA:** Bacteriophage lytic enzymes: novel anti−infectives. Trends Microbiol 2005, 13, 491–496.
- **[42] Borysowski J, Weber−Dabrowska B, Gorski A:** Bacteriophage endolysins as a novel class of antibacterial agents. Exp Biol Med 2006, 231, 366–377.
- **[43] Fischetti VA:** Novel method to control pathogenic bacteria on human mucous membranes. Ann N Y Acad Sci 2003, 987, 207–214.
- **[44] Loeffler J, Nelson D, Fischetti VA:** Rapid killing of *Streptococcus pneumoniae* with a bacteriophage hydrolase. Science 2001, 294, 2170–2172.
- **[45] Cheng Q, Nelson D, Zhu S, Fischetti VA:** Removal of group B streptococci colonizing the vagina and orophar− ynx of mice with a bacteriophage lytic enzyme. Antimicrob Agents Chemother 2005, 49, 111–117.
- **[46] Jado I, Lopez R, Garcia E, Fenoll A, Casal J, Garcia P:** Phage lytic enzymes as therapy for antibiotic−resistant *Streptococcus pneumoniae* infection in a murine sepsis model. J Antimicrob Chemother 2003, 52, 967–973.
- **[47] Loeffler J, Djurkovic S, Fischetti VA:** Phage lytic enzyme Cpl−1 as a novel antimicrobial for pneumococcal bac− teremia. Inf Immun 2003, 71, 6199–6204.

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Conflict of interest: The Institute has filed a patent application covering therapeutic use of phages in bacterial infections.

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